

Supporting Information for:

Selection in males purges the mutation load on female fitness

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Table S1: Comparison and rationale for using opposite-sex heterosis for point estimates and sex-averaged heterosis for sex-differences. Using opposite-sex heterosis is the most technically correct approach for the point estimates and assessment of whether they are different from zero, but causes male and female estimates to not be directly comparable. Using sex-averaged heterosis renders the sex-specific estimates directly comparable – and is therefore the most technically correct assessment of fold differences and significance between the sexes – but the estimates may be biased due to shared measurement error between fitness and heterosis. Estimates and values used to support the conclusions are bolded.

Description	Heterosis	Sex	Symbol	Estimate (CIs)	<i>P</i> value	Fold difference	Sex-diff. <i>P</i> value
Genetic standardized selection	Opposite-sex	Male	β'_{a_M}	-0.0125 (-0.031, -0.003)	0.008	8.7x	0.11
		Female	β'_{a_F}	-0.0014 (-0.013, 0.009)	0.672		
	Sex-averaged	Male	β''_{a_M}	-0.0133 (-0.029, -0.004)	0.008	3.7x	0.104
		Female	β''_{a_F}	-0.0036 (-0.011, 0.007)	0.558		
Genetic correlation	Opposite-sex	Male	$r_{o_M, o_F - i_F}$	-0.59 (-0.81, -0.11)	0.008	-	0.132
		Female	$r_{o_F, o_M - i_M}$	-0.14 (-0.40, 0.28)	0.672		
	Sex-averaged	Male	-	-0.46 (-0.79, -0.12)	0.008	-	0.12
		Female	-	-0.06 (-0.37, 0.17)	0.558		

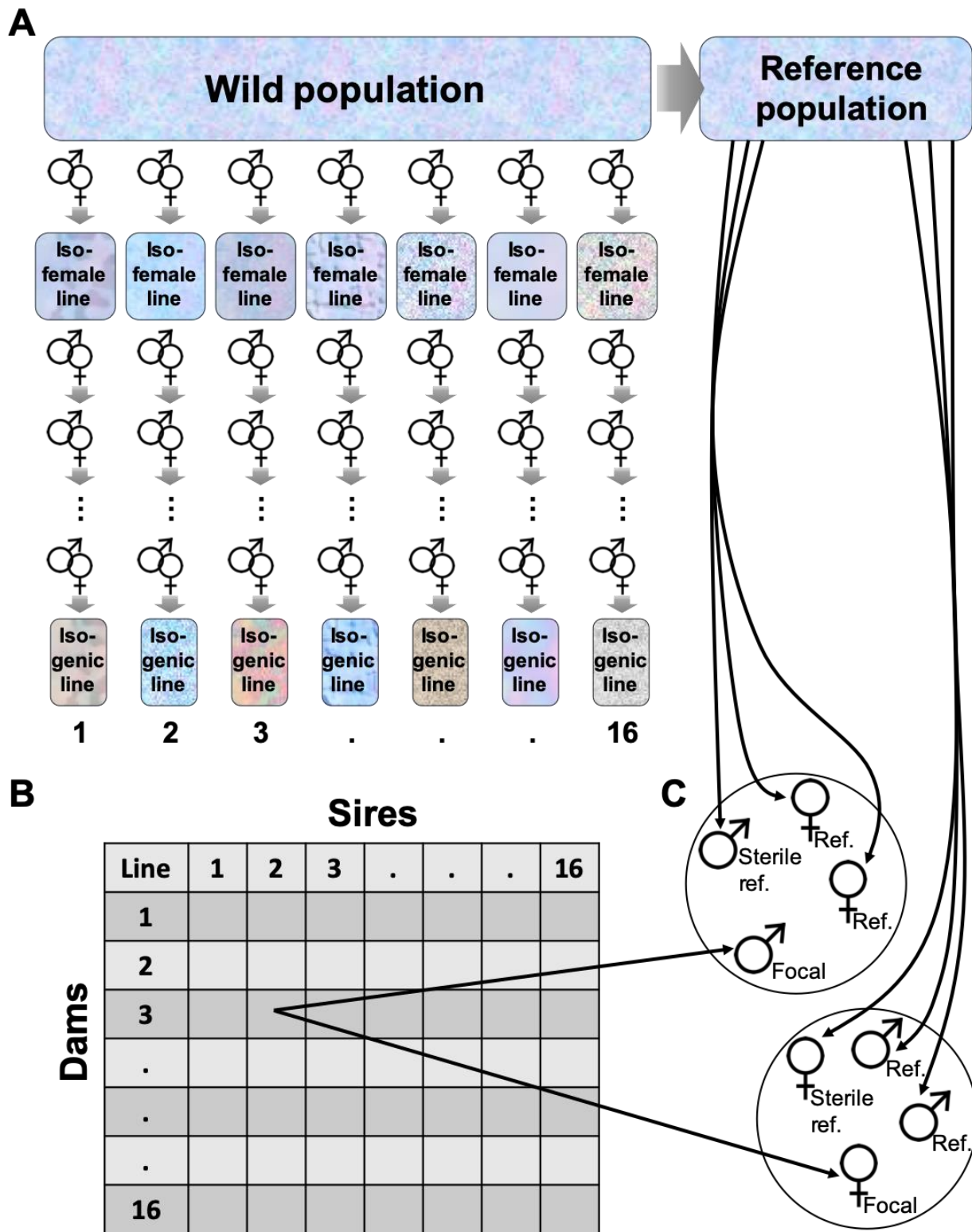


Fig. S1: Lome population history and experimental design. (A) A wild population was divided into an outbred laboratory reference population and 41 isofemale lines (Berger et al. 2014). The latter were further inbred (Grieshop et al. 2017) to obtain the 16 inbred/isogenic lines used presently (Grieshop and Arnqvist 2018). Genetic diversity is depicted by color and texture. (B) A full diallel cross (Lynch and Walsh 1998) among the 16 inbred strains, where F_1 inbred parental selfs (i) are on the diagonal and outcrossed F_1 s (o) are on the off-diagonal. (C) Replicate F_1 male and female fitness estimates included a focal F_1 individual (e.g. outbred (o) F_1 s from a strain-2 sire and strain-3 dam), a sterile same-sex competitor from the reference population, two (fertile) opposite-sex competitors from the reference population, and ca. 100 *V. unguiculate* seeds in a 90 mm \varnothing petri dish. These beetles were left to interact/mate/oviposit for the duration of their lifetime, and F_2 offspring counts = focal individuals' fitness.

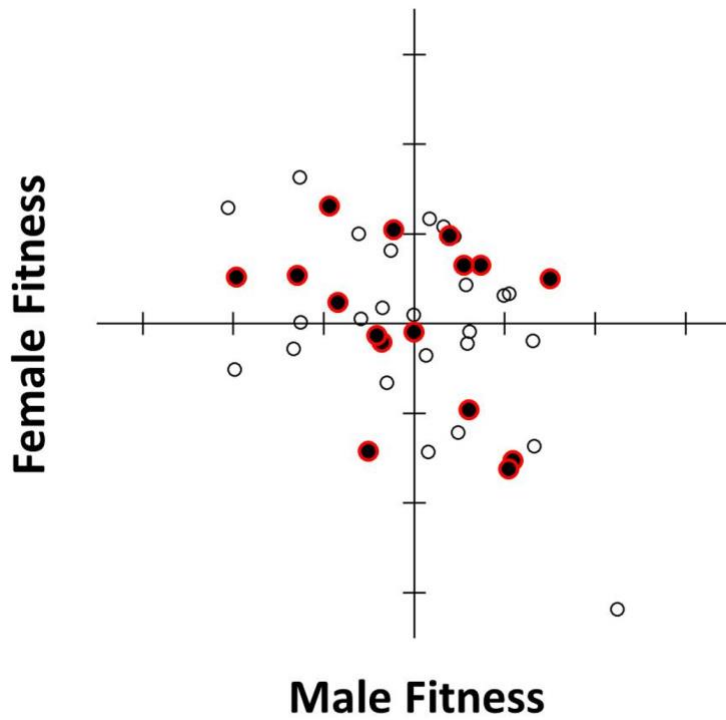


Fig. S2: The raw-means r_{MF} of the isofemale lines (see Fig. S1; Berger et al. 2014). Those from whence the present 16 inbred strains stem are outlined in red and filled in black. Because all 20 replicate inbreeding lineages stemming from some of the most male-benefit/female-detriment isofemale lines went extinct prior to completing the full inbreeding program (Grieshop et al. 2017), the present 16 inbred strains were chosen from throughout the isofemale line r_{MF} with the aim of countering that bias.

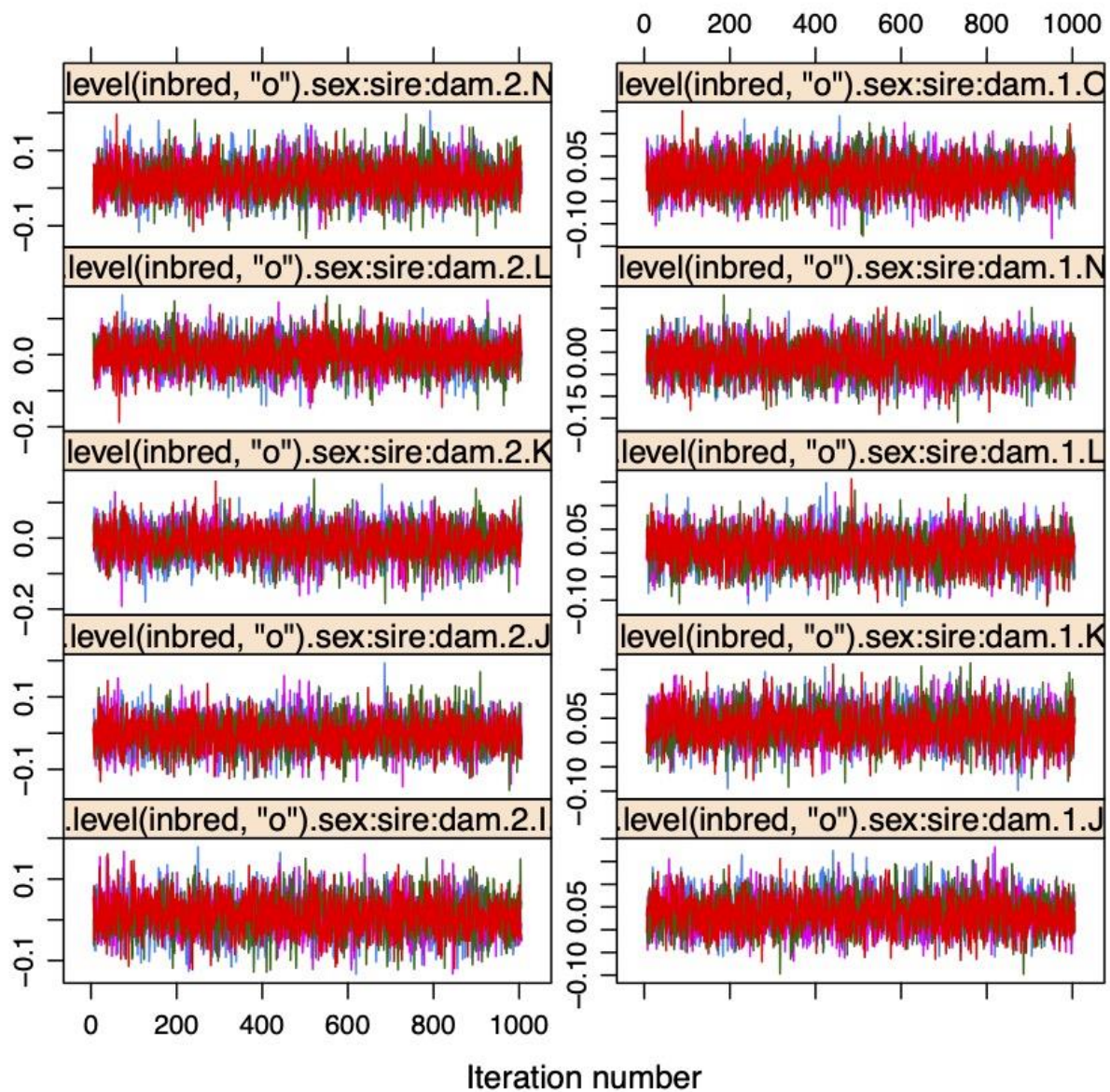


Fig. S3: Results the Gelman-Rubin analysis, demonstrating good mixing of four independent MCMC chains. Shown is a representative random sample of ten parameters, out of the total 780 parameters (plus an intercept) that were estimated in our model. Other diagnostic output from this analysis is available in the R script (starting on line 110).

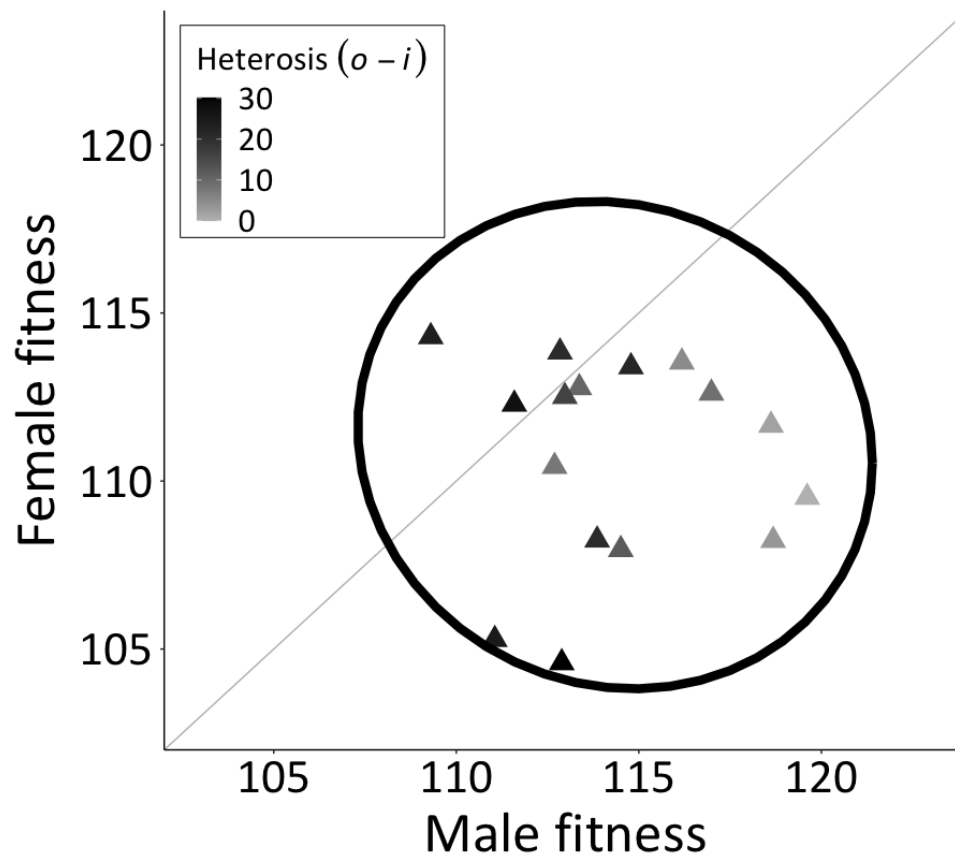


Fig. S4: Outbred (o) breeding values from Fig. 1A shaded by sex-averaged heterosis. An informal depiction of our main finding is apparent in that heterosis is clearly distributed along a horizontal gradient, according to male breeding values for fitness, but is clearly not distributed along the vertical dimension. Thus, sex-/strain-specific heterosis, the degree to which inbred strains benefit from having their rare partially recessive deleterious mutations covered up by heterozygosity, is reflected in those strains' outcrossed males, but not their outcrossed females.

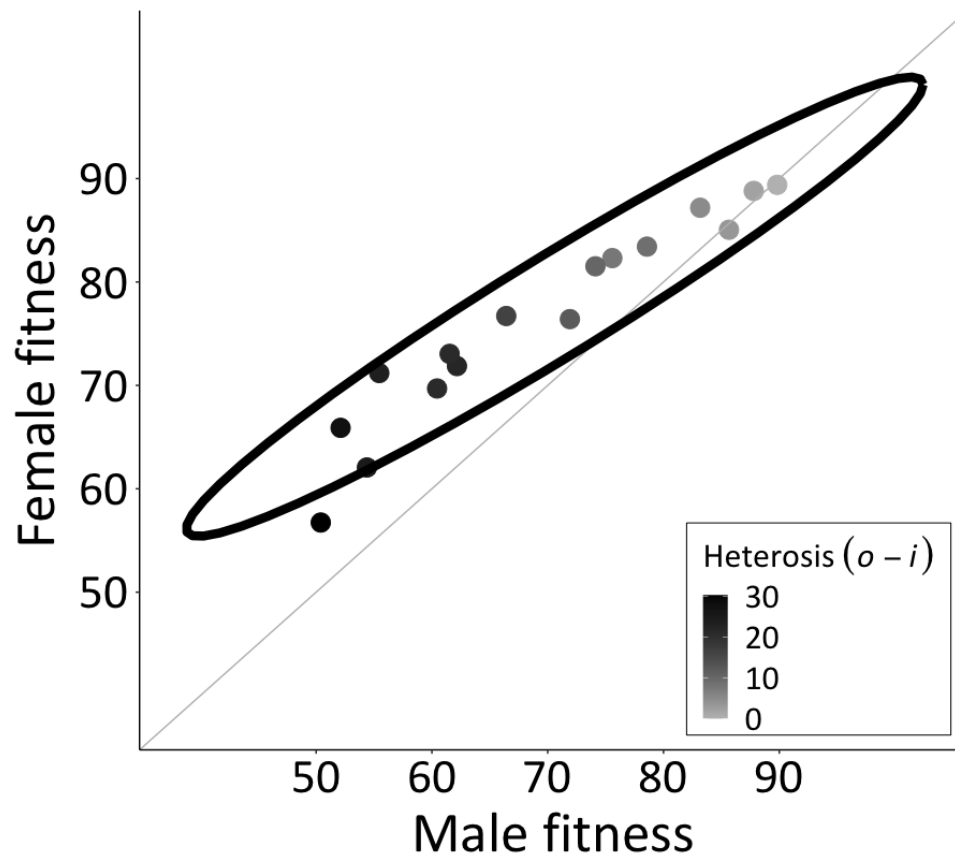


Fig. S5: Inbred (*i*) breeding values from Fig. 1A shaded by sex-averaged heterosis. By definition, strains with greater inbred fitness experience less heterosis. Male fitness is more negatively impacted by inbreeding/homozygosity than female fitness, as the large majority of these strains' inbred breeding values lie above the $y=x$ line.

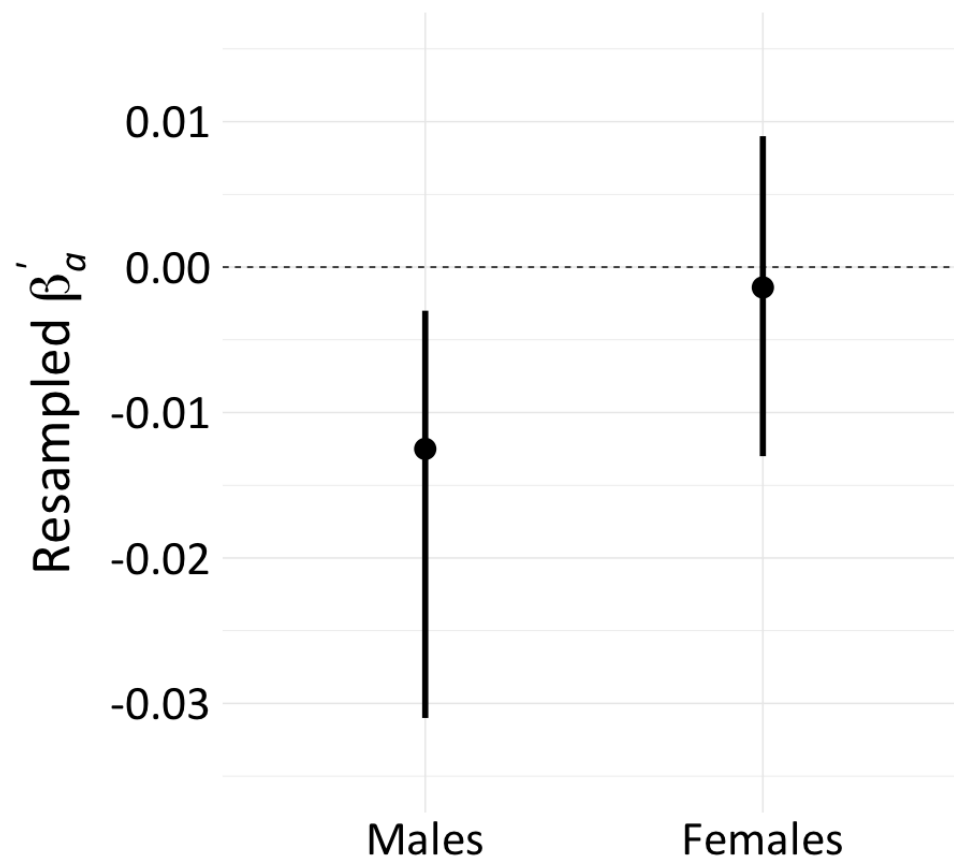


Fig. S6: Resampled point estimates and 95% credibility intervals of β'_a for males (β'_{a_M}) and females (β'_{a_F}).

SI References

Berger, D., Grieshop, K., Lind, M.I., Goenaga, J., Maklakov, A.A. and Arnqvist, G. (2014). Intralocus sexual conflict and environmental stress. *Evolution* 68:2184-2196.

Grieshop, K., Berger, D. and Arnqvist, G. (2017). Male-benefit sexually antagonistic genotypes show elevated vulnerability to inbreeding. *BMC Evol Biol* 17:134.

Grieshop, K. and Arnqvist, G. (2018). Sex-specific dominance reversal of genetic variation for fitness. *PLoS Biol* 16:e2006810.